

## Article

# Formulation Optimization of Extemporaneous Oral Liquids Containing Naloxone and Propranolol for Pediatric Use

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**Abstract:** There is a need to develop dosage forms suitable for children to improve drug treatment. Extemporaneous compounding of drugs for children is one way to meet these needs. However, excipients generally considered as safe in adults may not be appropriate in dosage forms intended for children. The aim was to optimize the composition of two pediatric liquid preparations by substituting paraben as a microbiological preservative and ethanol as a solubilizer, with excipients more suitable for pediatric use. The oral liquids were Naloxone 1 mg/mL and Propranolol 10 mg/mL. Twelve different formulations were tested with propranolol and naloxone, respectively, during the screening process to select appropriate formulations. Sodium benzoate and glycerol were used as a preservative and solubilizer, respectively, and different pH of the formulations were evaluated. The formulations were characterized according to dispensed dose (dosing accuracy), viscosity and osmolality. The optimized formulations from the screening process were tested with two amounts of sodium benzoate and microbiological assays were performed. These formulations were shown to have satisfactory preservative properties and dosing accuracy. The results showed that the oral liquids could be prepared without the addition of solubilizer and with lower osmolality (naloxone), thus reducing the risk of gastrointestinal side effects.

**Keywords:** extemporaneous compounding; oral liquid; preservative; solubilizer; pediatric use



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## 1. Introduction

Extemporaneous compounding of drugs for children is used in order to meet the need for drug formulations and doses suitable for children. Many drugs are administered orally and only available as solid oral dosage forms, mainly tablets. These may be unsuitable for children due to lack of appropriate doses and swallowing difficulties among young children. Furthermore, off-label use of medicine is a common practice in the pediatric population [1]. In addition, it may be difficult to divide a commercially available solid dosage form in a correct and reproducible manner when administering the medicine to children [2–4]. Modifications of solid oral dosage forms, such as tablet splitting, tablet crushing, and capsule opening can potentially affect therapeutic outcomes for patients and adverse events have been reported because of medicine modifications [5,6]. Consequently, there is a need for suitable pediatric formulations to improve drug treatment of children [7–12]. Oral suspensions and solutions may be an alternative to solid oral dosage forms and if no commercial liquid formulation is available, these can be manufactured extemporaneously. Oral liquid formulations can be rather easily manufactured extemporaneously, can provide flexibility regarding dosing and administration, and are easy to swallow [13–15]. One way of preparing extemporaneous oral liquids is to use commercially available solid oral dosage forms such as tablets and capsules, for example crushing of tablets or opening of capsules and adding water or another vehicle. When preparing an extemporaneous compound for pediatric use, the starting compound is often intended for adults and this needs to be taken

into consideration. One major concern regarding liquid formulations is their stability [16]. Microbiological preservatives are generally added to the formulation to prevent growth of microorganisms and improve stability and shelf life. Another issue that needs to be addressed is taste, thus ensuring a formulation that will be acceptable to the pediatric population [13–15]. Osmolality and viscosity are also important properties to take into consideration when formulating oral liquids. Viscosity may affect the acceptability and palatability of the formulation and dosing accuracy [17]. Osmolality should not exceed 700 mOsm/kg in oral liquids to minimize gastrointestinal side effects [18]. Choosing excipients suitable for children is another challenge when developing pediatric formulations since excipients generally considered safe in adults may not be appropriate in dosage forms intended for children [19,20].

There are several reasons to use microbiological preservatives in liquid formulations, e.g., to protect the user against microbial infections and to prevent chemical or physical changes caused by microorganisms, which in turn can affect the stability and the appearance of the formulation. Preservation of a liquid formulation can be achieved by altering the physicochemical properties, e.g., pH, of the formulation or by adding a chemical substance to prevent microbial growth [21]. According to literature, pH between 2 and 9 can be tolerated in oral solutions and pH of a pharmaceutical solution is a compromise between solubility and stability as well as excipient functionality and physiological acceptability [22].

The optimal microbiological preservative should be effective against a broad spectrum of microorganisms. Most microbiological preservatives are weak acids and bases and, consequently, the pH of the formulation will affect the microbial effect of the preservative [21]. Type and properties of the dosage form and safety issues also need to be taken into account when choosing a suitable preservative. Safety issues are of particular importance in the pediatric population. Properties such as adequate solubility, an acceptable taste and smell, compatibility with active pharmaceutical ingredients (API), excipients and packaging, environmentally safety, broad antimicrobial spectra, low toxicity, effective in low concentrations, and costs need to be considered when choosing an appropriate preservative [21]. However, a single preservative does not possess all these properties; in some cases, it is a matter of choosing the least unsuitable preservative.

Some excipients, such as propylene glycol, ethanol, sorbitol and parabens, are considered potentially harmful in children [19]. In a recent publication, Rouaz et al. critically reviewed the use of excipients in the pediatric population and emphasized the need for safe and appropriate excipients for children [20]. In addition, Zupanets et al. compiled information of excipients used in pediatric cough syrups and concluded that most of the investigated formulations contained excipients potentially harmful to the pediatric population [23]. The safety of excipients used especially in infants, where metabolic and elimination pathways are not yet fully developed and who are more susceptible to adverse effects, must be taken into consideration [24]. Therefore, the selection of appropriate excipients in pediatric formulations is challenging. Parabens such as methyl parahydroxybenzoate (methylparaben) and propyl parahydroxybenzoate (propylparaben) are used as microbiological preservatives in drug formulations and in extemporaneous compounding of oral liquids. Parabens are regarded as “generally recognized as safe” (GRAS) by the Food and Drug Administration (FDA) in the USA [25]. Concern has been raised regarding the use of parabens in drug formulations regarding their negative effects on the endocrine system [26]. For example, in 2015 European Medical Agency (EMA) published a reflection paper regarding the use of parabens as excipients in oral formulations [27]. At present, there are not enough clinical data to support a change of recommendations of their use, but a limitation on their use in children could be warranted. In the STEP (Safety and Toxicity of Excipients for Paediatrics) database, information regarding excipients and their use in pediatrics is compiled and this information can be used to facilitate the selection of appropriate excipients [28]. Some studies have shown that parabens may be unsuitable for children. One study conducted kinetic modelling based on blood samples from newborns and showed that a daily exposure of drugs containing methylparaben can cause prolonged

exposure due to unmetabolized methylparaben [29]. Another study showed that there was a risk of hyperbilirubinemia in newborns, since methylparaben can bind to albumin and thus displace bilirubin [30]. This risk was also observed with propylparaben in children younger than six months [31].

Ethanol is used as a solvent in oral liquid preparations. In extemporaneous compounding of oral liquids, a stock solution of paraben in ethanol is often used due to the low aqueous solubility of paraben and to facilitate manufacturing. Treating, for example, chronic conditions in children with drug formulations containing ethanol may lead to acute and chronic intoxication, and ethanol can affect both absorption and metabolism of drugs when administered at the same time [9,24]. One study showed that children treated with dosage forms containing ethanol in some cases obtained amounts exceeding half of the safe weekly consumption in adult women [32]. Another study showed that infants and newborns can be exposed to amounts of ethanol exceeding 6 mg/kg/day during drug treatment [33]. Regular exposure (weekly) of ethanol in small amounts is contraindicated for children younger than six years [9].

Two different oral liquids were included in the present study: Naloxone 1 mg/mL and Propranolol 10 mg/mL. These oral liquids are extemporaneously manufactured by Apotek Produktion & Laboratorier AB (APL) in Sweden. The choice of these two preparations was based on use (i.e., pediatric use), composition (i.e., original composition containing paraben and ethanol), and production volume (i.e., high production volume). Naloxone is used in the treatment of opioid-induced constipation and propranolol is used in the treatment of arrhythmia and hypertension. The physicochemical properties of naloxone and propranolol are presented in Table 1.

**Table 1.** Physicochemical properties of the active pharmaceutical ingredients used in the liquid formulations [34–37].

| Property                                      | Naloxone   | Propranolol |
|---|------------|-------------|
| pH stability (drug)                           | 2.5–5.0    | 2.8–4.0     |
| pH stability (product)                        | 3.0–4.0    | ≤3          |
| pKa   | 7.84–10.07 | 9.42        |
| Water solubility (drug) (mg/mL)               | 5.64       | 0.079       |
| Water solubility (hydrochloride salt) (mg/mL) | 50         | 10          |

The aim of the study was to optimize the composition of two pediatric extemporaneous liquid preparations containing naloxone and propranolol by substituting paraben as a microbiological preservative and ethanol as a solubilizer, with excipients more suitable for pediatric use. The present study is a response to concerns raised by the European Medicines Agency (EMA), regarding safety for children when using parabens. Due to the lack of clinical data, EMA has not yet changed their recommendation for paraben use in liquid formulations, although some clinical studies indicate that parabens are not suitable for pediatric use.

## 2. Materials and Methods

### 2.1. Materials

Two different oral liquids were chosen to be included in the study: Naloxone 1 mg/mL and Propranolol 10 mg/mL. These preparations are provided by APL in Sweden. An overview of the materials and their function in the oral liquids is given in Table 2. The original compositions of the two oral liquids are presented in Table 3.

**Table 2.** An overview of the materials and their functions. API = Active pharmaceutical ingredient.

| Material   | Function   | Manufacturer                             | Country                     |
|--|--|--|-----------------------------|
| Naloxone hydrochloride dihydrate   | API  | Sanofi Chimie                            | France                      |
| Propranolol hydrochloride  | API  | Cosma S.p.A.                             | Italy                       |
| Stock solution of methyl parahydroxybenzoate<br>Ethanol 96%<br>Methyl parahydroxybenzoate<br>Water, purified | Preservative   | Univar<br>Merck KGaA<br>APL <sup>1</sup> | Sweden<br>Germany<br>Sweden |
| Sodium benzoate  | Preservative   | Emerald Performance Materials            | Netherlands                 |
| Sodium citrate   | Buffering agent  | Jungbunzlauer AG                         | Austria                     |
| Citric acid monohydrate  | Buffering agent  | Jungbunzlauer AG                         | Austria                     |
| Sugar syrup 63%<br>Sucrose<br>Methyl parahydroxybenzoate<br>Water, purified                                  | Sweetening agent,<br>viscosity-increasing agent          | Nordic Sugar AB<br>Merck KGaA<br>APL     | Sweden<br>Germany<br>Sweden |
| Glycerol 85%<br>Glycerol<br>Water, purified  | Solvent, sweetening agent,<br>viscosity-increasing agent | Oleon NV                                 | France                      |
| Hydrochloric acid 1 M and 5 M  | pH adjustment  | Merck KGaA                               | Germany                     |
| Water, purified  | Solvent  | APL                                      | Sweden                      |

<sup>1</sup> APL = Apotek Produktion & Laboratorier AB.**Table 3.** Original composition of the two oral liquids included in the study.

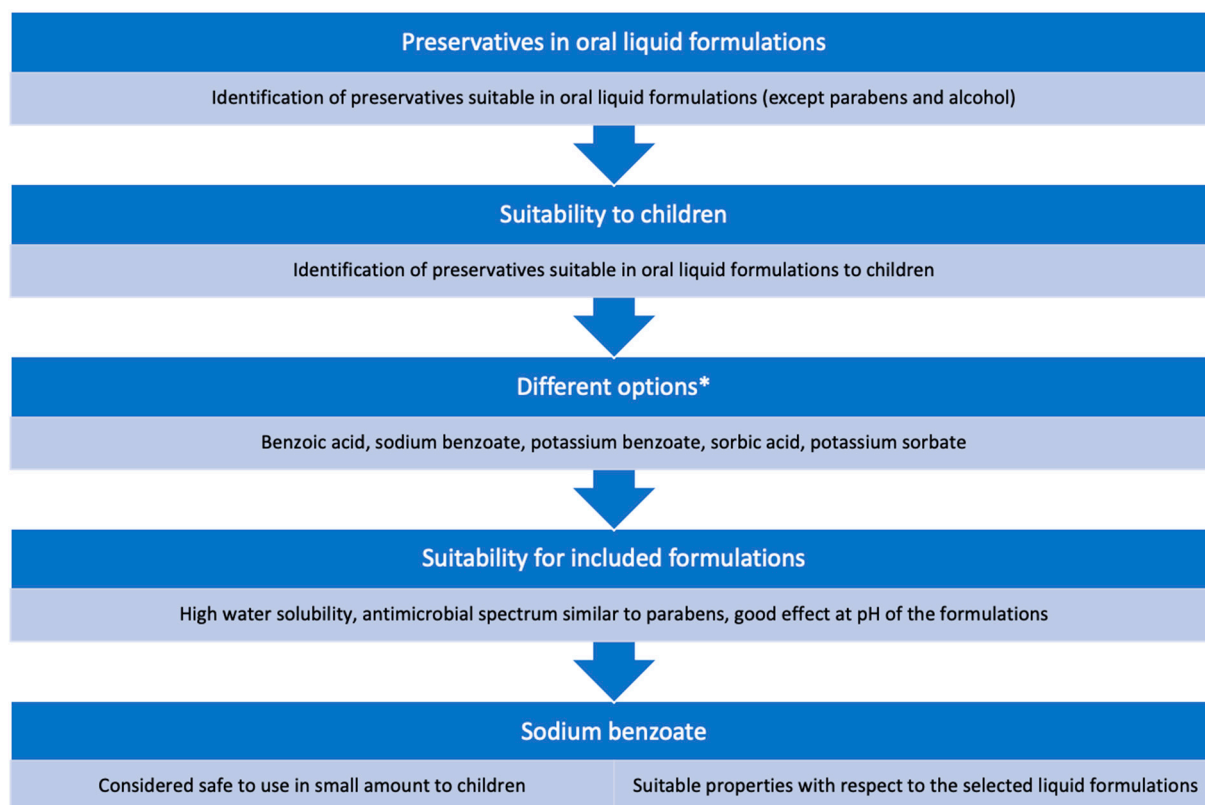
| Preparation                    | Material  | Amount        |
|--------------------------------|---|---------------|
| Naloxone 1 mg/mL               | Naloxone hydrochloride dihydrate                                  | 1 mg          |
|                                | Methyl parahydroxybenzoate <sup>1</sup><br>(+6 mg Ethanol 96%)    | 1 mg          |
|                                | Hydrochloric acid 1 M   | q.s. pH 3–3.5 |
|                                | Water, purified   | ad 1 mL       |
| Propranolol 10 mg/mL<br>pH 3.9 | Propranolol hydrochloride   | 10 mg         |
|                                | Sodium citrate  | 2.1 mg        |
|                                | Citric acid monohydrate   | 2.8 mg        |
|                                | Sugar syrup   | 320 mg        |
|                                | Methyl parahydroxybenzoate <sup>1</sup><br>(+4.08 mg Ethanol 96%) | 0.68 mg       |
|                                | Water, purified   | ad 1 mL       |

<sup>1</sup> Methyl parahydroxybenzoate is used as stock solution containing 10% methyl parahydroxybenzoate (composition per 100 g: methyl parahydroxybenzoate 10 g, ethanol (96%) 60 g, purified water 30 g). ad = to; q.s. (quantum satis) = the amount which is enough.

## 2.2. Choice of Preservative and Solubilizer

There are several microbiological preservatives that are suitable for liquid formulations. Figure 1 shows a flow chart used to select an appropriate preservative to substitute methyl parahydroxybenzoate in liquid formulations intended for children. The following properties of the microbiological preservative were considered important: high water solubility, antimicrobial spectra similar to parabens, and good antimicrobial effects at the pH of the liquid formulations. Considering these different aspects of a preservative, sodium benzoate was chosen as the microbiological preservative [28,38–40]. In the original compositions, ethanol was primarily used as a solubilizer for the preservative and a stock solution of the preservative was used. Excluding ethanol in the formulation could poten-

tially affect the solubility of the API and therefore an alternative solublizer was evaluated. Ethanol (0.4–0.6%) was substituted by glycerol (0–20%) in the formulations since glycerol has previously been shown to be well tolerated by children [41–43].



**Figure 1.** Flow chart describing the process to select an appropriate preservative in order to substitute methyl parahydroxybenzoate. \* Benzoic acid: solubility in water at 20 °C 1 in 300, optimal antimicrobial pH 2.5–4.5, effective against bacteria, fungi, mold and yeast. Sodium benzoate: solubility in water at 20 °C 1 in 1.8, optimal antimicrobial pH 2–5, effective against bacteria and fungi. Potassium benzoate: solubility in water at 20 °C 1 in 2.36, optimal antimicrobial pH ≤ 4.5, effective against bacteria and fungi. Sorbic acid: insoluble in water, optimal antimicrobial pH 4.5, effective primarily against fungi. Potassium sorbate: solubility in water at 20 °C 1 in 1.72, optimal antimicrobial pH < 6, effective primarily against fungi [8,22,28,38–40,44].

### 2.3. Manufacture of the Liquid Formulations

The liquid formulations were manufactured according to internal protocol at APL in Sweden. 50 mL was generally prepared. However, in the challenge tests, where the preservative effect was analyzed, a larger volume was required, and 500 mL was therefore manufactured in these tests. The excipients and API were first weighed. Glycerol (when included) and sugar syrup (when included) were first added to a volumetric flask and thereafter 80% of the stipulated amount of water. The remaining dry excipients were added one at a time while stirring. The pH was then adjusted, and the solution was diluted with water to the final volume. The original liquid formulations were manufactured and characterized regarding dispensed dose (dosing accuracy), osmolality and viscosity, and they were used as a baseline to compare with the corresponding characteristics of the modified formulations.

A small experiment was first performed to investigate the compatibility with the new preservative, sodium benzoate, and to obtain pH values of the formulations to be used in the subsequent design of experiments. The liquid formulations were prepared using sodium benzoate instead of methyl parahydroxybenzoate; the other ingredients were the same as in the original formulations. The concentration of sodium benzoate was set to 0.5%,

which is the highest recommended amount according to the Handbook of Pharmaceutical Excipients [44]. However, in the formulation containing propranolol, the concentration of sodium benzoate had to be reduced to 0.2% since a precipitate was observed, probably caused by the preservative. This precipitation was observed at 0.3% sodium benzoate.

#### 2.4. Experimental Plan

To study the effect of substituting methyl parahydroxybenzoate and ethanol with sodium benzoate and glycerol more systematically, a design of experiment (DoE) approach was used. An experimental plan for the experiments was developed using MODDE Pro (Sartorius Stedim Biotech, Sweden). Variables included in the experiments were pH and amount of glycerol (%). The pH interval was selected based on the results of the screening experiments and the stability of each API (Table 1). These variables were then combined according to the principle high/low, low/high, low/low and high/high. Tables 4 and 5 show the experimental plans for the liquid formulations.

**Table 4.** Design of experiments for Naloxone 1 mg/mL. Name and order of experiments, pH to be set, and the amount of solubilizer (%). N5, N10–12 are replicates. The amount of sodium benzoate was 0.5%.

| Order of Manufacturing | Experiment Number | Experiment Name | pH   | Glycerol (%) |
|------------------------|-------------------|-----------------|------|--------------|
| 1                      | 7                 | N7              | 3    | 20           |
| 2                      | 4                 | N4              | 3    | 10           |
| 3                      | 3                 | N3              | 4.5  | 0            |
| 4                      | 6                 | N6              | 4.5  | 10           |
| 5                      | 2                 | N2              | 3.75 | 0            |
| 6                      | 1                 | N1              | 3    | 0            |
| 7                      | 9                 | N9              | 4.5  | 20           |
| 8                      | 12                | N12             | 3.75 | 10           |
| 9                      | 11                | N11             | 3.75 | 10           |
| 10                     | 8                 | N8              | 3.75 | 20           |
| 11                     | 5                 | N5              | 3.75 | 10           |
| 12                     | 10                | N10             | 3.75 | 10           |

**Table 5.** Design of experiments for Propranolol 10 mg/mL. Name and order of experiments, pH to be set, and the amount of solubilizer (%). N5, N10–12 are replicates. Amount of sodium benzoate was 0.2%.

| Order of Manufacturing | Experiment Number | Experiment Name | pH | Glycerol (%) |
|------------------------|-------------------|-----------------|----|--------------|
| 9                      | 9                 | N9              | 5  | 20           |
| 3                      | 3                 | N3              | 5  | 0            |
| 10                     | 10                | N10             | 4  | 10           |
| 5                      | 5                 | N5              | 4  | 10           |
| 12                     | 12                | N12             | 4  | 10           |
| 8                      | 8                 | N8              | 4  | 20           |
| 2                      | 2                 | N2              | 4  | 0            |
| 7                      | 7                 | N7              | 3  | 20           |
| 11                     | 11                | N11             | 4  | 10           |
| 1                      | 1                 | N1              | 3  | 0            |
| 6                      | 6                 | N6              | 5  | 10           |
| 4                      | 4                 | N4              | 3  | 10           |

#### 2.5. Optimization and Evaluation Using MODDE Pro

The following responses were used in the study; viscosity (mean value at 20 s<sup>−1</sup>), solubility (1 = soluble, 0 = insoluble) and dispensed dose. These responses were included in MODDE Pro when developing the design of experiments. A response contour plot was



obtained for each liquid formulation. The variables pH and amount (%) glycerol were also included. Based on the response contour plots, new compositions of optimal formulations were obtained. Liquid formulations based on these optimal formulations and with varied amounts of preservative were then manufactured and analyzed thereafter with respect to dispensed dose, viscosity, osmolality and preservative effect.

## 2.6. Evaluation of Oral Liquids

### 2.6.1. Solubility

Solubility of the API in the oral liquids was determined visually by ocular inspection of the vials after manufacturing.

### 2.6.2. pH

The pH of the oral liquids was determined using a pH meter (Mettler Toledo Seven Multi, Mettler Toledo, Columbus, OH, USA). Hydrochloric acid (1 M or 5 M) or sodium hydroxide (0.1 M or 1 M) were added to adjust pH to the specified value. Measurements were conducted according to European Pharmacopoeia (Ph. Eur.) 2.2.3—Potentiometric determination of pH [45].

### 2.6.3. Viscosity

The viscosity of the oral liquids was determined using a rheometer (Discovery HR-1, TA Instruments, UK). Measurements were performed using a cone-plate, geometry 6 cm with a cone angle of 1°. A temperature of 20 °C, linear mode, and shear rate from 3 to 110 s<sup>−1</sup> were used. Measurements were conducted according to Ph. Eur. 2.2.10—Viscosity—rotating viscometer method) [45]. Measurements were performed in duplicate. The results are presented as mean values ± coefficient of variation.

### 2.6.4. Osmolality

The osmolality of the oral liquids was measured using an osmometer (Mikro-Osmometer Automatic, Löser Messtechnik, Berlin, Germany). The osmometer was calibrated according to Ph.Eur. 2.2.35—Osmolality [45] using 100 µL standard solution with known osmolality (300 mOsm/kg H<sub>2</sub>O) (300 standard solution for osmometer (Löser Messtechnik, Germany)) instead of using Water R as specified in Ph.Eur. Measurements were conducted in duplicate. The results are presented as mean values ± coefficient of variation.

### 2.6.5. Dispensed Dose and Dosing Accuracy

Dosing accuracy of the oral liquids was tested to evaluate the suitability of the liquid formulations together with the dosing device. A suitable dose was selected for each formulation based on use in children. For Naloxone 1 mg/mL, the dose was set at 1 mg (1 mL) based on doses previously used in pediatric patients [46]. Dropleas adapters and dosing syringes (5 mL) were used to withdraw the doses. For Propranolol 10 mg/mL, the dose was set at 5 mg (0.5 mL) based on a dose of 1–5 mg/kg bodyweight/day and assuming a bodyweight of 5 kg and dosing three times a day. Dropleas adapters and dosing syringes (1 mL) were used to withdraw the doses. All doses were weighed on an analytical balance (Mettler Toledo XP603S, Mettler Toledo, USA). Density measurements of the solutions were performed using a density meter (DMA 4100 M, Anton Paar, Graz, Austria). The volume of the dose withdrawn by the syringe (dispensed dose) was used as a measure of dosing accuracy of the liquid formulations. The volume was calculated by dividing the weight of the dose with the density of the solution (n = 10). The results are presented as mean value ± 95% confidence interval. An acceptable dosing accuracy was set to ±10% of target dose (volume) (according to internal guidelines at APL).

### 2.6.6. Preservative Effect

The preservative effect was evaluated using a challenge test for 28 days according to Ph.Eur. 5.1.3—Efficacy of antimicrobial preservation [45]. During the test, the oral liquids

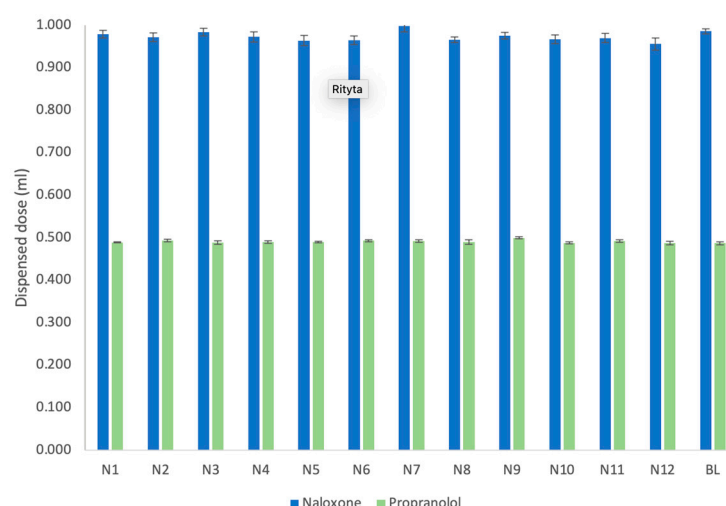
were challenged with different microorganisms and the liquids were stored at a prescribed temperature (20–25 °C). At specified time intervals, samples were withdrawn from the liquids and the number of microorganisms in the samples was counted. Two formulations containing naloxone and propranolol, respectively, were chosen. The formulations were chosen based on the evaluation in MODDE Pro. Two different amounts of preservative were tested (0.05% and 0.1% *w/v* sodium benzoate).

### 3. Results and Discussion

Twelve different formulations (N1–N12) were tested with propranolol and naloxone, respectively, during the screening process to be able to select appropriate formulations (Tables 4 and 5). The formulations were characterized according to dispensed dose (dosing accuracy), viscosity and osmolality. The results were compared with the formulations with the original compositions (baseline).

#### 3.1. Dispensed Dose and Dosing Accuracy

Figure 2 shows dispensed doses of the different formulations containing naloxone and propranolol, respectively, in comparison with the baseline formulations. The results showed that all formulations had an acceptable dosing accuracy and that there were small differences between the formulations. For naloxone formulations (N1–N12), the dosing accuracy, i.e., error in dispensed dose, varied between 0.2% and 4.4% and for propranolol formulations (N1–N12); the error in dispensed dose varied between 0% and 2.4%.



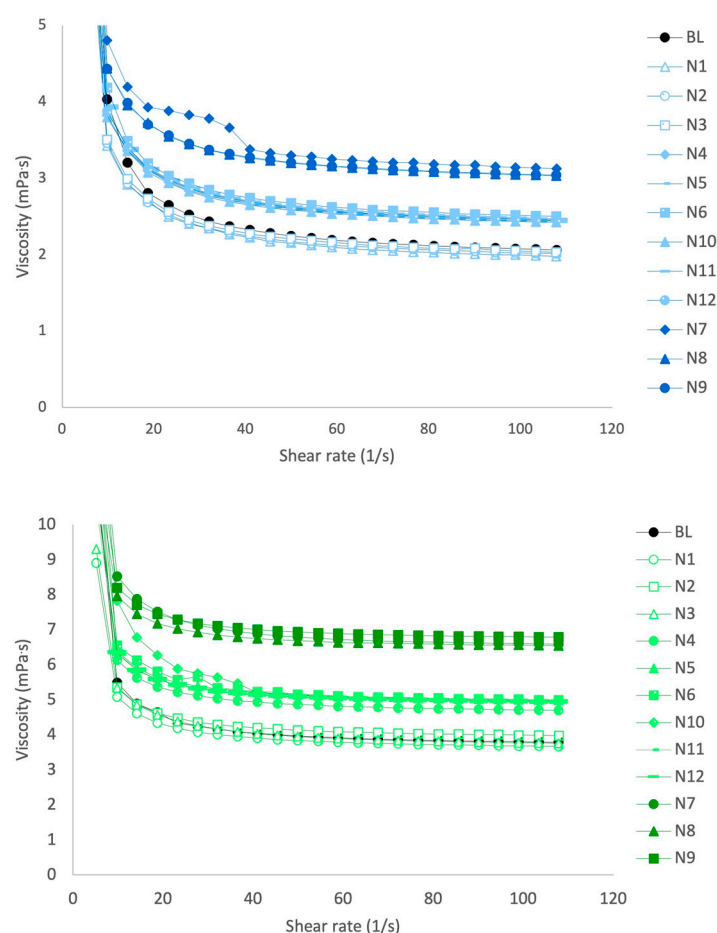
**Figure 2.** Dispensed dose (dosing accuracy) of formulations containing naloxone (blue) and propranolol (green). BL = baseline (original composition). Compositions of formulations N1–N12 according to Tables 4 and 5. Mean value with 95% confidence interval ( $n = 10$ ). The target dose was 1 mL (corresponding to 1 mg naloxone) and 0.5 mL (corresponding to 5 mg propranolol).  $\pm 10\%$  of the target dose was considered an acceptable dosing accuracy.

The type of dosing device can affect dosing accuracy and studies have shown that using a syringe is generally better than a cup, dropper or spoon to accurately dispense a dose [47–50] but the suitability of a dosing device was also shown to be dependent on the viscosity of the liquid [47]. Furthermore, dosing accuracy is also related to consumer use and education of the consumer on the correct way to use a dosing device [47–50]. A high dosing accuracy of the formulation together with a, for the formulation, suitable dosing device is crucial in order ensure that a correct dose is administered [51,52]. The differences in dosing accuracy between the oral liquids containing naloxone and propranolol may be due to differences in syringes used (5 mL vs. 1 mL syringe). Higher dosing accuracy (lower dispensing error) was generally obtained with Propranolol 1 mg/mL in which the smaller syringe was used.



### 3.2. Viscosity

The results regarding viscosity are presented in Figure 3. The viscosity of all formulations decreased with an increase in shear rate. The viscosity was affected by the amount of glycerol in the formulations and the viscosity increased with increased amounts of glycerol, which was expected due to the higher extent of hydrogen bonding in glycerol compared with water. For the oral liquids with the original composition (baseline), the viscosity was similar to the formulations containing 0% glycerol. The formulations with propranolol had a higher viscosity compared with the formulations with naloxone, which is probably due to the presence of sugar syrup in the propranolol formulations. Sugar syrup is used as a sweetener in oral liquids but also as a viscosity-increasing agent.



**Figure 3.** Viscosity as a function of shear rate for the oral liquids containing naloxone ((**top figure**), blue) and propranolol ((**bottom figure**), green) (mean value,  $n = 2$ ). BL = baseline (original composition). Compositions of formulations N1–N12 according to Tables 4 and 5.

Viscosity has been reported to affect dosing accuracy, especially when using a cup or dropper compared with a syringe [47,49]. Higher viscosity of the oral liquid has been shown to decrease dosing accuracy which may potentially affect the administered dose [47,49]. Viscosity may also affect the acceptability and palatability of the formulation [17]. Although there were some differences in viscosity between the investigated formulations, all formulations obtained acceptable dosing accuracy. A higher viscosity did not have a negative impact on dosing accuracy in the present study, which may be due to a relatively low viscosity of the formulations and the relatively small difference in viscosity between the formulations.

### 3.3. Solubility

Naloxone was insoluble in the oral liquids with pH 3 (N1, N4 and N7) and soluble in the oral liquids with pH 3.75 and 4.5. Propranolol was insoluble in the oral liquids with pH 5 and 0% and 10% glycerol (N3 and N6) and soluble in the oral liquids with pH 5 and 20% glycerol and all oral liquids with pH 3 and 4. Apparently, the solubility of both naloxone and propranolol is pH dependent, and a suitable pH of the formulation must be chosen. The addition of glycerol did not generally have an effect on solubility, suggesting that pH is more important for the solubility of the drug substances compared with addition of a cosolvent/solubilizer. The observations indicated that it was possible to obtain solutions of both naloxone and propranolol given the right circumstances. Solubility was only determined visually which may have affected the interpretations due to the subjectivity of the method. In extemporaneous compounding, solubility is often determined by visual inspection. However, alternative methods to determine solubility, such as the shake-flask method, could provide a more objective analysis of solubility and is recommended in future studies.

### 3.4. Osmolality

Hyperosmolar oral liquids have been shown to cause gastrointestinal side effects such as feeding intolerance and delayed gastric emptying [18]. Osmolality should not exceed 700 mOsm/kg in oral liquids in order to minimize gastrointestinal side effects [18]. In newborns, osmolality around 400 mOsm/kg is desirable [53,54] and a recent review concluded that feed osmolality of 300–500 mOsm would not likely pose a safety risk to newborn infants [55]. However, a recent study showed that the osmolality of a majority of commonly used oral medication in neonates was greater than 500 mOsm/kg [56]. Thus, osmolality is an important property to take into consideration prior to administration of oral liquids. Osmolality could only be measured with the oral liquid containing propranolol and 0% glycerol since the osmometer could only measure osmolarities up to 2500 mOsm/kg (Table 6). The oral liquids containing propranolol and 10% and 20% glycerol had an osmolality exceeding 2500 mOsm/kg. Osmolality of all the oral liquids containing naloxone could be measured. To be able to compare with propranolol, only osmolality for the oral liquids containing 0% glycerol is presented in Table 6. The osmolality of formulations containing propranolol was higher than in formulations containing naloxone. Adding glycerol to the formulations containing naloxone resulted in increased osmolality (data not shown).

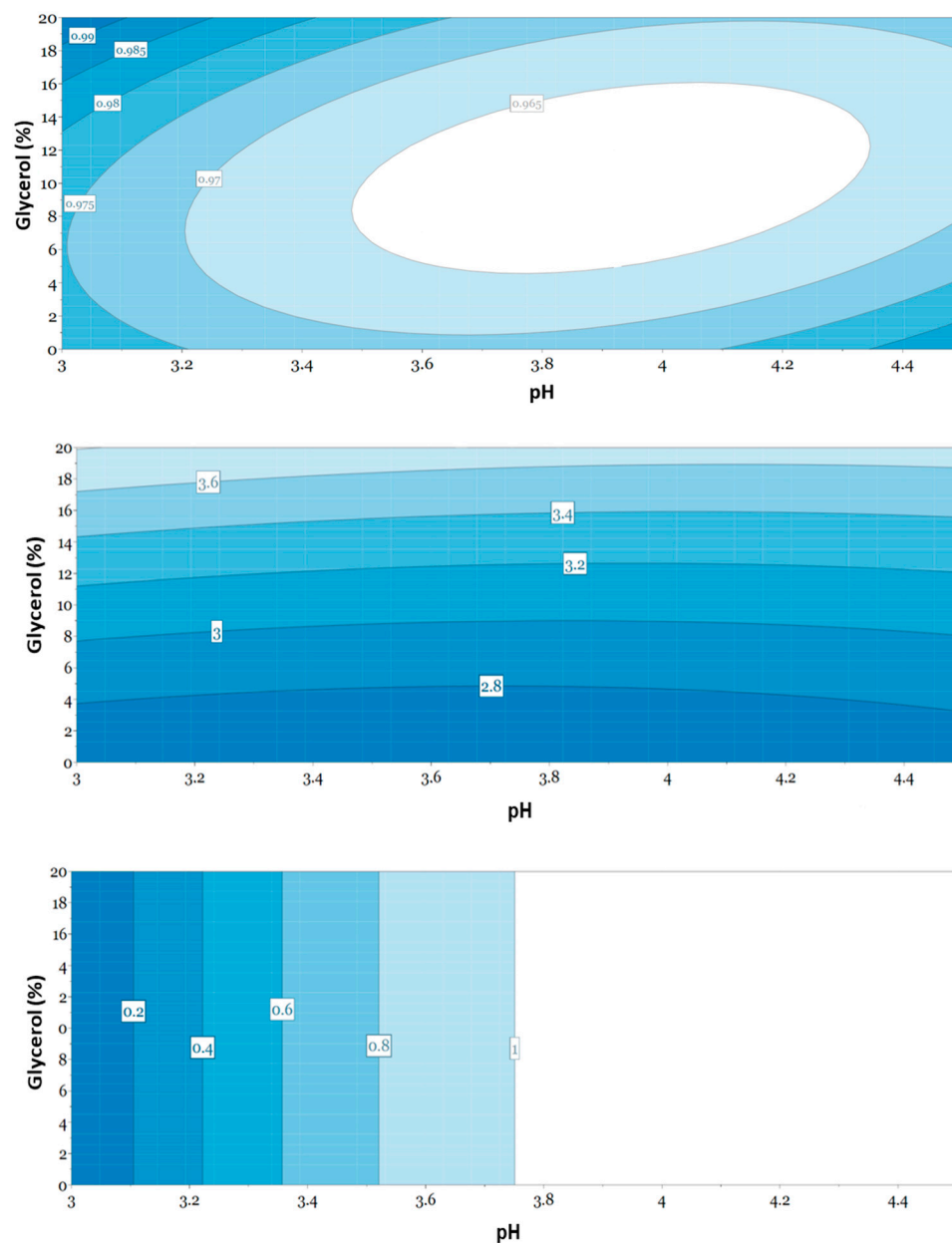
**Table 6.** Osmolality of the oral liquids containing naloxone and propranolol with 0% glycerol. Mean value  $\pm$  coefficient of variation ( $n = 2$ ). Compositions of formulations N1–N3 according to Tables 4 and 5.

| Naloxone 1 mg/mL | Osmolality (mOsm/kg) | Propranolol 10 mg/mL | Osmolality (mOsm/kg) |
|------------------|----------------------|----------------------|----------------------|
| Baseline         | 136 $\pm$ 0.007      | Baseline             | 983 $\pm$ 0.005      |
| N1               | 97 $\pm$ 0.000       | N1                   | 1121 $\pm$ 0.011     |
| N2               | 96 $\pm$ 0.000       | N2                   | 941 $\pm$ 0.011      |
| N3               | 81 $\pm$ 0.000       | N3                   | 918 $\pm$ 0.003      |

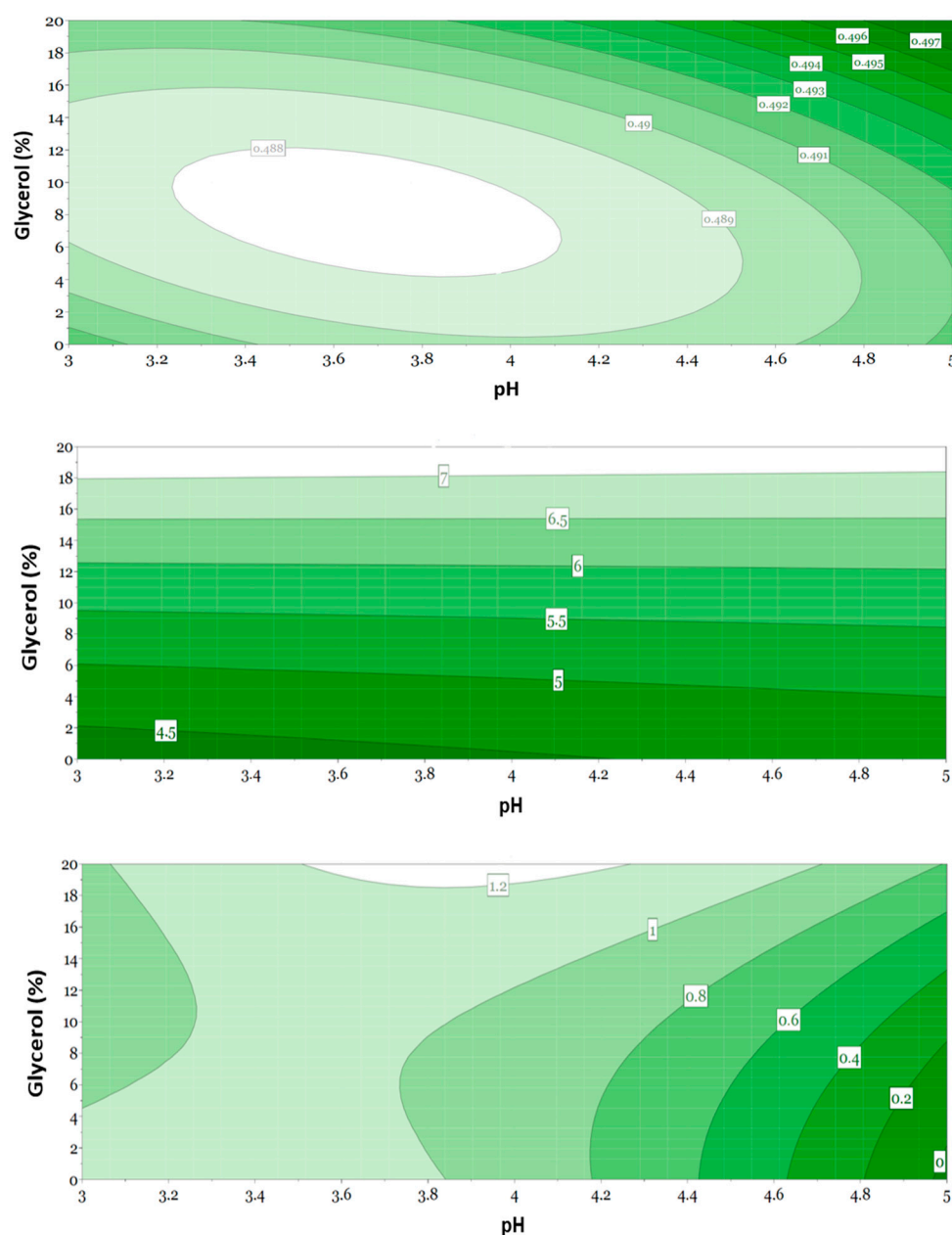
### 3.5. Evaluation of Responses Using Multivariate Data Analysis

To understand the effect of the selected variables on the selected responses, response contour plots were created using multivariate data analysis. The responses included were viscosity (mean value at 20 s<sup>−1</sup>), solubility (1 = soluble, 0 = insoluble) and dispensed dose and the variables were pH and amount of glycerol (%). With a DoE approach, it is possible to minimize the number of experiments and still gain valuable insights regarding the effect of formulation variables on the quality of the formulations. The goal is to develop an optimized formulation for the selected APIs by using multivariate data analysis. Figures 4 and 5 show the response contour plots for the oral liquids containing naloxone

and propranolol, respectively. The results with naloxone showed that the best dispensed dose (dosing accuracy) can be obtained using compositions with either 0% or 20% glycerol and pH 3 or compositions with 0% glycerol and pH 4.45 (Figure 4). Viscosity increased with increasing amounts of glycerol and was independent of pH of the formulation (Figure 4). For naloxone to be soluble in the formulation, pH should be set to 3.75 or higher. Solubility of naloxone was independent of the amount of glycerol (Figure 4).



**Figure 4.** Response contour plot for responses dispensed dose (dosing accuracy) (**top figure**), viscosity (**middle figure**) and solubility (**bottom figure**) as a function of amount of glycerol and pH. Compositions of formulations N1–N12 according to Table 4, containing naloxone. The target dose was 1 mL, corresponding to 1 mg naloxone.  $\pm 10\%$  of the intended dose was considered an acceptable dosing accuracy. Values above 1 in the solubility plot indicate that naloxone is soluble in the formulation, values below 1 indicate that naloxone is not soluble.



**Figure 5.** Response contour plot for responses dosing accuracy (dispensed dose) (**top figure**), viscosity (**middle figure**) and solubility (**bottom figure**) as a function of amount of glycerol and pH. Compositions of formulations N1–N12 according to Table 5, containing propranolol. The target dose was 0.5 mL, corresponding to 5 mg propranolol.  $\pm 10\%$  of the target dose was considered an acceptable dosing accuracy. Values above 1 in the solubility plot indicate that propranolol is soluble in the formulation, values below 1 indicate that propranolol is not soluble.

The results with propranolol showed that the best dispensed dose (dosing accuracy) was obtained with compositions with pH 5 and 20% glycerol (Figure 5). Using 0% glycerol, the best dosing accuracy was obtained with compositions with pH 3. Viscosity increased with increasing amount of glycerol and was independent of pH (Figure 5). For propranolol to be soluble in the formulations, pH should be set to 3–3.85 in compositions containing 0% glycerol (Figure 5).

### 3.6. Evaluation of Selected Formulations

Based on how the variables pH and amount of glycerol affected the responses solubility, dispensed dose and viscosity, optimal compositions were selected for naloxone and

propranolol, respectively (Table 7). This selection was based on the interpretation of the contour plots, as described above, to identify a suitable amount of glycerol and suitable pH to be able to optimize the solubility, dispensed dose and viscosity of the liquids. When selecting the composition, the stability of API at different pH (Table 1) and the optimal pH of the selected preservative sodium benzoate (Figure 1) were also taken into consideration. Based on the results from the screening process, glycerol was chosen not to be included as a solubilizer because solubility of the API could be achieved without the addition of glycerol and glycerol increased the osmolality. The pH had an effect on solubility of the API and consequently, the pH of the oral liquids has to be optimized and an acceptable pH-interval was therefore suggested (Table 7). pH in both the original and the optimized formulations is rather low. The literature suggests that pH 2–9 can be tolerated in oral liquids [22] and studies have shown that oral liquids for pediatric use exhibit a wide range of pH, from acidic to basic [57,58]. Previous studies have shown that pH, as well as sweeteners, may cause erosive damage to the enamel surface of the teeth [57] and pH of oral liquids is therefore an important aspect to consider.

**Table 7.** Selected compositions of the two oral liquids after evaluation using multivariate data analysis.

| Preparation          | Material                         | Amount          |
|----------------------|----------------------------------|-----------------|
| Naloxone 1 mg/mL     | Naloxone hydrochloride dihydrate | 1 mg            |
|                      | Sodium benzoate <sup>1</sup>     | 0.5 or 1 mg     |
|                      | Hydrochloric acid 1 M            | q.s. pH 3.9–4.1 |
|                      | Water, purified                  | ad 1 mL         |
| Propranolol 10 mg/mL | Propranolol hydrochloride        | 10 mg           |
|                      | Sodium citrate                   | 2.1 mg          |
|                      | Citric acid monohydrate          | 2.8 mg          |
|                      | Sugar syrup                      | 320 mg          |
|                      | Sodium benzoate <sup>1</sup>     | 0.5 or 1 mg     |
|                      | Hydrochloric acid 1 M            | q.s. pH 3.2–3.6 |
|                      | Water, purified                  | ad 1 mL         |

<sup>1</sup> 0.05% and 0.1% sodium benzoate were tested. ad = to; q.s. (quantum satis) = the amount which is enough.

The selected (optimized) compositions with two different amounts of sodium benzoate were characterized regarding dispensed dose (dosing accuracy), viscosity, osmolality (Table 8) and preservative effect (Tables 9 and 10). The results showed that dosing accuracy was acceptable for all selected formulations when dispensed using a syringe (Table 8). In the present study, drug content in each dose was not measured but it is suggested in future optimization studies to measure drug content with an appropriate analytical method. The viscosity and osmolality of the formulations appeared to be lower compared with the original compositions. However, since the measurements were made in duplicate, a more thorough statistical analysis is not possible, and interpretations should be made with caution.

As mentioned earlier, osmolality should not exceed 700 mOsm/kg in oral liquids to minimize gastrointestinal side effects [18]. The optimized formulations with naloxone all showed osmolalities well below 700 mOsm/kg. However, the optimized formulations with propranolol had osmolalities above 700 mOsm/kg. Despite this, gastrointestinal side effects have not been reported with Propranolol 10 mg/mL (original composition), suggesting a safe pediatric use also for the modified formulations. However, in future optimization studies, the high osmolality of the propranolol formulations should be addressed.

**Table 8.** Characterization of the selected (optimized) compositions regarding dispensed dose (dosing accuracy), viscosity and osmolality. Mean value  $\pm$  95% confidence interval of dispensed dose (% error) (n = 10), mean value  $\pm$  coefficient of variation of viscosity and osmolality (n = 2). Naloxone dose = 1 mL (corresponding to 1 mg naloxone) and propranolol dose = 0.5 mL (corresponding to 5 mg propranolol). Acceptable dosing accuracy (i.e., error in dispensed dose) was set to  $\pm$ 10% of target dose.

| Preparation          | Dispensed Dose (mL) (% Error) | Viscosity <sup>1</sup> (mPa·s) | Osmolality (mOsm/kg) |
|----------------------|-------------------------------|--------------------------------|----------------------|
| Naloxone 1 mg/mL     |                               |                                |                      |
| Baseline             | 0.986 $\pm$ 0.006 (1.4)       | 2.72 $\pm$ 0.017               | 136 $\pm$ 0.007      |
| SB 0.05%             | 0.959 $\pm$ 0.008 (4.1)       | 2.60 $\pm$ 0.008               | 13 $\pm$ 0.000       |
| SB 0.1%              | 0.972 $\pm$ 0.006 (2.8)       | 2.61 $\pm$ 0.005               | 22 $\pm$ 0.000       |
| Propranolol 10 mg/mL |                               |                                |                      |
| Baseline             | 0.487 $\pm$ 0.004 (2.6)       | 4.48 $\pm$ 0.016               | 983 $\pm$ 0.005      |
| SB 0.05%             | 0.491 $\pm$ 0.001 (1.8)       | 4.22 $\pm$ 0.012               | 933 $\pm$ 0.002      |
| SB 0.1%              | 0.491 $\pm$ 0.001 (1.8)       | 4.31 $\pm$ 0.015               | 952 $\pm$ 0.003      |

SB = sodium benzoate, AV = acceptance value, Baseline = original composition with methylparaben. <sup>1</sup> At shear rate 20 1/s.

**Table 9.** Microbiological challenge test for Naloxone 1 mg/mL with preservative sodium benzoate. Results expressed as log<sub>10</sub> reduction after 14 and 28 days.

| Microorganism                   | Amount Sodium Benzoate (%) | Start CFU/mL                 | Log <sub>10</sub> Reduction 14 Days | Log <sub>10</sub> Reduction 28 Days |
|---------------------------------|----------------------------|------------------------------|-------------------------------------|-------------------------------------|
| <i>Staphylococcus aureus</i>    | 0.05                       | 2.4 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 2.4 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Pseudomonas aeruginosa</i>   | 0.05                       | 2.8 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 2.8 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Escherichia coli</i>         | 0.05                       | 3.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 3.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Candida albicans</i>         | 0.05                       | 6.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 6.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Aspergillus brasiliensis</i> | 0.05                       | 5.3 $\times$ 10 <sup>5</sup> | 2                                   | NI                                  |
|                                 | 0.1                        | 5.3 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |

*Staphylococcus aureus* (ATCC 6538), *Pseudomonas aeruginosa* (ATCC 9027), *Escherichia coli* (ATCC 8739), *Candida albicans* (ATCC 10231), *Aspergillus brasiliensis* (ATCC 16404) ATCC = American Type Culture Collection accession number; CFU = Colony Forming Unit; NI = No increase in number of viable microorganisms compared to the previous reading.

**Table 10.** Microbiological challenge test for Propranolol 10 mg/mL with preservative sodium benzoate. Results expressed as log<sub>10</sub> reduction after 14 and 28 days.

| Microorganism                   | Amount Sodium Benzoate (%) | Start CFU/mL                 | Log <sub>10</sub> Reduction 14 Days | Log <sub>10</sub> Reduction 28 Days |
|---------------------------------|----------------------------|------------------------------|-------------------------------------|-------------------------------------|
| <i>Staphylococcus aureus</i>    | 0.05                       | 2.4 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 2.4 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Pseudomonas aeruginosa</i>   | 0.05                       | 2.8 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 2.8 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Escherichia coli</i>         | 0.05                       | 3.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 3.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Candida albicans</i>         | 0.05                       | 6.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 6.5 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
| <i>Aspergillus brasiliensis</i> | 0.05                       | 5.3 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |
|                                 | 0.1                        | 5.3 $\times$ 10 <sup>5</sup> | 5                                   | NI                                  |

*Staphylococcus aureus* (ATCC 6538), *Pseudomonas aeruginosa* (ATCC 9027), *Escherichia coli* (ATCC 8739), *Candida albicans* (ATCC 10231), *Aspergillus brasiliensis* (ATCC 16404) ATCC = American Type Culture Collection accession number; CFU = Colony Forming Unit; NI = No increase in number of viable microorganisms compared to the previous reading.



Results from the microbiological challenge tests are shown in Tables 9 and 10. For bacteria, a  $\log_{10}$  reduction of at least 3, and for fungi, a  $\log_{10}$  reduction of at least 1, are required after 14 days, and the number of viable microorganisms must not have increased after 28 days compared to the value at 14 days to show successful preservation [45]. All formulations with naloxone and propranolol passed the microbiological challenge test (Tables 9 and 10). The formulations with propranolol also contained sugar syrup which may contribute to the preservative effect since sugar has a preservative effect of its own. In addition, the sugar syrup used contains 0.1% methylparaben. Although the oral liquid with propranolol contains some methylparaben, it was chosen to be included in the study to be able to compare two different products and to investigate if it was possible to prepare the liquids in a similar way although the API differed. To totally exclude parabens from the propranolol formulation, sugar syrup should be removed or replaced. To increase the acceptability of the product among children, some other sweetening or flavoring agent could be considered, such as xylitol or sucrose. Another alternative is to use sugar syrup with a higher concentration (85%) since this sugar syrup does not require any preservative. This is also applicable to the naloxone formulations since these does not contain any flavoring or sweetening agent. In the further development and optimization of the formulations, the overall acceptability and suitability of sweetening agents must be considered. There are safety issues related to use of sweeteners in drug formulations to pediatric patients, for example cariogenic and laxative effects [20].

There are no formal requirements to test extemporaneous formulations with respect to preservative effects since the shelf life of these products is short. Based on the stability data of up to 28 days storage and the rule of thumb that preservation should be conducted with as little preservative as possible, the lower amount of sodium benzoate, i.e., 0.05%, is probably sufficient. The amount of sodium benzoate generally used in pharmaceutical products is 0.02–0.5% [40]. The effect of the preservative may decrease with time due to degradation and chemical instability, resulting in a decrease in the amount of preservative in the dosage form. Concerns have been raised regarding the safety of sodium benzoate in formulations intended for children [59]. Benzoic acid and benzoates could displace bilirubin from albumin thus increasing the risk of hyperbilirubinemia. This is especially of concern in neonates younger than eight weeks where metabolic enzymes are not fully matured [60]. The acceptable daily intake of benzoic acid and benzoates has been set to 0–5 mg/kg body weight [61,62]. In the formulations suggested in the present study, the amount of sodium benzoate is well below the acceptable daily intake based on pediatric dosing.

#### 4. Conclusions

New compositions were suggested for Naloxone 1 mg/mL and Propranolol 10 mg/mL where methyl parahydroxybenzoate was replaced by sodium benzoate as preservative. The oral liquids containing sodium benzoate were shown to have satisfactory preservative properties during storage for up to 28 days. Glycerol was evaluated as a substitute for ethanol as a solubilizer but was not considered necessary to include in the new compositions since solubility of naloxone and propranolol could be achieved without addition of glycerol and glycerol increased osmolality. Furthermore, the oral liquids had an acceptable dosing accuracy. pH was rather low but within the tolerable range for oral liquids. In future formulation optimization studies of these oral liquids, aspects such as pH, osmolality and taste should be further addressed.

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