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Potential Upstream Regulators of Cannabinoid Receptor 1 Signaling in Prostate Cancer: A Bayesian Network Analysis of Data From a Tissue Microarray

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BACKGROUND. The endocannabinoid system regulates cancer cell proliferation, and in prostate cancer a high cannabinoid CB1 receptor expression is associated with a poor prognosis. Down-stream mediators of CB1 receptor signaling in prostate cancer are known, but information on potential upstream regulators is lacking.

RESULTS. Data from a well-characterized tumor tissue microarray were used for a Bayesian network analysis using the max-min hill-climbing method. In non-malignant tissue samples, a directionality of pEGFR (the phosphorylated form of the epidermal growth factor receptor) → CB1 receptors were found regardless as to whether the endocannabinoid metabolizing enzyme fatty acid amide hydrolase (FAAH) was included as a parameter. A similar result was found in the tumor tissue, but only when FAAH was included in the analysis. A second regulatory pathway, from the growth factor receptor ErbB2 → FAAH was also identified in the tumor samples. Transfection of AT1 prostate cancer cells with CB1 receptors induced a sensitivity to the growth-inhibiting effects of the CB receptor agonist CP55,940. The sensitivity was not dependent upon the level of receptor expression. Thus a high CB1 receptor expression alone does not drive the cells towards a survival phenotype in the presence of a CB receptor agonist.

CONCLUSIONS. The data identify two potential regulators of the endocannabinoid system in prostate cancer and allow the construction of a model of a dysregulated endocannabinoid signaling network in this tumor. Further studies should be designed to test the veracity of the predictions of the network analysis in prostate cancer and other solid tumors. Prostate 74: 1107–1117, 2014. © 2014 The Authors. The Prostate published by Wiley Periodicals, Inc. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

KEY WORDS: prostate cancer; epidermal growth factor; cannabinoid receptor; fatty acid amide hydrolase; directed acyclic graph

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INTRODUCTION

In 1975, Munson and colleagues reported that Δ⁶-tetrahydrocannabinol, the main psychoactive constituent of cannabis, reduced tumor growth, and increased life span in a xenograft model of lung cancer [1]. Since then, a large number of studies have demonstrated that the endocannabinoid system, defined here as cannabinoid CB₁ and CB₂ receptors, their endogenous ligands anandamide (AEA) and 2-arachidonoylglycerol (2-AG), and their synthetic and metabolic enzymes, control cancer cell proliferation, and migration [2–5]. The prostate gland expresses a functional epithelial CB₁ receptor, which in the rat is involved in the control of the contraction of the gland [6,7], and its expression is increased in prostate cancer (Pca) cells [8,9], as is the expression of several endocannabinoid metabolizing enzymes, such as fatty acid amide hydrolase (FAAH) [10,11], cyclooxygenase-2 [12], and acylglycerol kinase [13]. Data from cell cultures and animal models are consistent with the hypothesis that the circulating endocannabinoid system can dampen the proliferation and invasivity of Pca cells [14–17], although mitogenic effects of low concentrations of CB₁ receptor agonists secondary to activation of the Akt signaling pathway have been described [18]. In Pca tumor samples, CB₁ receptor scores and pAkt scores are correlated and show a significant interaction term in ordinal regression analyses with the Gleason score as the dependent variable [19].

Whilst down-stream mediators of CB₁ receptor signaling in Pca have been well-investigated (review, see [4]), little is known concerning potential upstream regulators of this receptor. One way of identifying potential candidates is to undertake a Bayesian network analysis of available tissue microarray data, to prioritize potential upstream regulators of this receptor. One way of identifying potential upstream regulators of this receptor. One way of identifying potential upstream regulators of this receptor. One way of identifying potential upstream regulators of this receptor. One way of identifying potential upstream regulators of this receptor.

MATERIALS AND METHODS

Patient Material and Immunohistochemistry

The tissue material (formalin-fixed, paraffin-embedded samples) was collected at the Regional Hospital, Västerås, Sweden, between 1975 and 1991 from a total of 419 patients consecutively diagnosed with Pca following transurethral resection for lower urinary tract symptoms [23,24]. The patients had not received anticancer therapy before undergoing the transurethral resection procedure. The distribution of Gleason scores (number [%] of cases) in the material was 4–5, 98 (23%); 6, 110 (26%); 7, 72 (17%); 8–10, 139 (33%). The tumor stage (when given in the database) was la–1b, 210 (51%); 2, 107 (26%); 3, 81 (20%); 4, 13 (3%). A total of 48 (15%) of 322 cases were found to have metastases (bone scan) at diagnosis. The patients were followed until 2003 (for further details, see [23] [for the first 305 cases] and [24] [for the entire dataset]). Tissue microarrays were constructed and in general between 1 and 8 cores (usually 5, including both primary and secondary Gleason grade areas; tumor tissue) could be scored for the parameter in question. The mean % of the specimen that was tumor associated (% ca) varied from 5% to 100%, with cases >5% showing a poorer prognosis than those ≤5% [23]. The number (%) of cases at % ca levels of 5, 10–25, 30–50, 55–75, and 80–100, were 81 (19%), 125 (30%), 56 (13%), 46 (11%), and 111 (26%), respectively. Immunoreactive scores for the epithelial CB₁ receptor (CB₁R), EGFR, pEGFR, ErbB2, LRIG1, platelet-derived growth factor receptor β (PDGFRβ), androgen receptor (AR), von Willebrand factor (vWf), endoglin, hyaluronan, mast cell, and FAAH used in the present study were taken from our database (see [11,19,25–32] for published data and details of the methodologies). For the main parameters used here with the exception of LRIG1, the cores were scored on the basis of intensity and distribution to give a composite value. Thus, for example, a core scored at 25% intensity 1, 25% intensity 2, and 50% intensity 3 for the biomarker in question would score 0.25×1+0.25×2+0.5×3=2.25. The intensity ranges for CB₁R and ErbB2 were 0–4, pEGFR 0–5, LRIG1 0–2, and FAAH 0–3. The composite score thus represents a measure of the “loading” of the tumor for the parameter in question. For some parameters (e.g., FAAH [11]), the scores were made upon microscopic evaluation of the cores. This was also the case for the CB₁ receptors in our initial study using an intensity range of 0–3 [33]. However, we rescored digital images of the samples using a range 0–4 and two investigators, who returned scores with a very high inter-rater reliability (Chronbach’s alpha of 0.94 for an intra-class correlation analysis using a mixed model and testing for consistency [19]) and we used these values here. In both these examples, the investigators were blind to the clinical data, other than the obvious morphological characteristics of the samples, when scoring the cores. The antibodies used for the main parameters were: CB₁R, AbCam cat. no. 23703, lot no. 280229; pEGFR, Cell Signaling TYR845
lot 4; FAAH, an antibody raised against the last 102 amino acids of rat FAAH and kindly provided by Prof. Ken Mackie, Indiana University, USA; LRIG1, Agrisera rabbit polyclonal LRIG1-151; ErbB2, mouse c-erbB-2 Prediluted Cocktail Antibody, Biocare ([11,27,30,33] and unpublished data). The Ventana system was used for high-throughput immunostaining of the samples (see the above papers for details).

The research ethical committee at Umea university hospital (Regional Ethical Review Board in Umea, Sweden) approved of the study and waived the need for informed consent. In the database used for the analyses, the tissue samples were given a case number and year, and the patient names were not indicated in the database.

**CB₁ Receptor-Transfected AT1 Cells**

R3327-AT1 rat Dunning prostate cancer cells (ATCC, Manassas, VA; hereafter called AT1 cells) were cultured in RPMI medium (10% FBS, 1% penicillin-streptomycin, 2 mM glutamine, 250 mM dexamethasone). The cells were seeded out at 250,000–300,000 cells per well in 6-well plates. The next day, the cells were transfected either with the control plasmid (pIRES2-eGFP) or the murine CB₁ receptor-containing plasmid (pIRES2-mCB1-eGFP) using the TransIT™-prostate transfection kit and protocol supplied by the Mirus Corporation (Madison, WI). For details of the plasmids, see [34]. Initial experiments indicated that the ratio of DNA: TransIT™-reagent: Prostate Boost reagent (supplied in the kit) of 3 μg: 10 μl: 10 μl gave the best transfections. After incubation with the prostate boost reagent for 20 min, chloroquine (25 μM final concentration) was added and the samples were incubated for 150 min. Thereafter, the transfection media was replaced by media containing 10% (v/v) glycerol, the cells were incubated for 3 min at room temperature followed by two washes with warm phosphate-buffered saline. Finally, the culture media was added and the cells were allowed to grow for 48–72 hr prior to assessing the number of eGFP-positive cells by FACS analysis. Cells were selected in growth medium containing G418 (400 μg/ml). This protocol provided cells with a very large range of fluorescence intensity on FACS. Initial experiments indicated that incubation with a low concentration of CP55,940 ((–)-cis-3-[2-hydroxy-4-(1,1-dimethylheptyl)phenyl]-trans-4-(3-hydroxypropyl)cyclohexanol; Tocris Cookson, Bristol, UK), resulted in a loss of the highest intensity cells for both the eGFP- and CB₁/eGFP-transfected cells, suggesting that at this level of transfection, the plasmid load is detrimental to cell survival regardless of the absence or presence of the murine CB₁ receptors. In consequence, these cells were removed by a 6-day incubation of both eGFP- and CB₁/eGFP-transfected cells with 10 nM CP55,940 after which the cells were cultured for 2 weeks to amplify the stocks. In the experiments reported here, the cells, in six-well culture plates, were then incubated with test compounds for 3 days, and cell proliferation and fluorescence intensities were determined by FACS using a Guava easyCyte™ Flow Cytometer (Merck Millipore).

**Statistics**

Three statistical software programmes were used. Two-way ANOVA and Spearman’s correlation coefficients were determined using the statistical package built into the GraphPad Prism 5 and 6 computer programmes for the Macintosh (GraphPad Software Inc., San Diego, CA). Univariate regressions using the general linear model were undertaken using SPSS software (IBM SPSS statistics version 22 for the Macintosh, IBM Corporation, Armonk, NY). The directed acyclic graphs and bootstrap analyses were calculated using the function mmhc in the bnlearn package of the R computer programme [35].

**RESULTS**

**Interconnection between CB₁R, pEGFR, ErbB2, LRIG1, and FAAH in prostate tumor tissue.**

To identify potential components of a network that encompasses CB₁ receptors and is involved in Pca cell tumourigenesis, we undertook a simple bivariate correlation analysis with a number of different biochemical markers that have been scored in a well-characterized Pca tumor microarray (see [11,19,23–33] for hitherto published data). Using a cut-off Spearman’s rho value of 0.2, four parameters were identified: pEGFR, FAAH (as reported previously, [11,32]), the growth factor receptor ErbB2, and the EGFR regulatory protein LRIG1 (leucine-rich and immunoglobulin-like domains protein 1) (Fig. 1). These associations were not seen in the non-malignant tissue (Fig. 1).

Although the sample set were consecutive cases, the nature of their accruement (diagnosis following transurethral resection) means that they are not an unselected set of all Pca cases, since cases with Pca but not in the resected region will have been missed. Further, there has been a shift over time in the severity of the disease (i.e., relative incidence of the different tumor stages and Gleason scores) at diagnosis since the introduction of PSA testing [36]. In order to gain information as to whether this could be a limiting factor in our study, we conducted univariate regressions using the general linear model with the tumor CB₁ receptor scores as the dependent variables, the
Gleason scores (grouped as 4–6, 7, and 8–10) and the tumor stages (grouped as 1a–1b, 2, 3, and 4) as fixed factors, and each of the four parameters identified above as co-variates. In each case, the co-variates contributed significantly to the regression. The unstandardized beta weights (with 95% confidence intervals in brackets) for the four co-variates were: pEGFR, 0.125 (0.039–0.210), \( P < 0.005 \); FAAH, 0.224 (0.091–0.356), \( P < 0.001 \); ErbB2, 0.212 (0.117–0.306), \( P < 0.0001 \); LRIG1, 0.345 (0.161–0.530), \( P < 0.0005 \). In all cases, the Levene’s test of equality of error variances was satisfied (\( P > 0.3 \)). This indicates that the associations between the four parameters and the CB\(_1\) receptor score in the tumor tissue remain even when controlled for the severity of the disease at diagnosis.

For the non-malignant samples, the data available in the database was used to construct two directed acyclic graphs, one with three variables (CB\(_1\)R, luminal epithelial pEGFR and LRIG1), and one with four variables (CB\(_1\)R, luminal epithelial pEGFR, LRIG1, and luminal epithelial FAAH; non-malignant ErbB2 was not scored in a sufficient number of cores to be used). The method used to conduct the analysis was the max-min hill-climbing algorithm, which gives every possible network a score, and outlines a network with the highest score that best fits the data [37]. Further, the datasets were subjected to a bootstrap analysis and the process was repeated 1,000 times to estimate the uncertainty in the analyses. A bootstrap dataset of the same size as the original dataset is obtained by random sampling with replacement from the observations in the original data. For each bootstrap sample, a directed acyclic graph was constructed, and then the frequency of the possible combinations was obtained. This information was then incorporated in the directed acyclic graphs for the original complete dataset where the thickness of the edges indicates the reliability of the measure. The method requires that, for a given sample, there are no missing values. Thus, for example, if a network comprising three parameters is constructed, then every case must have scores for the parameters in question. This means that the optimal network size will be a trade-off between the number of parameters comprising the network and the number of cases scored for all the parameters. Sample sizes used here are given in the figure legends.

For the three-variable analysis, an edge pEGFR → CB\(_1\)R was found for the non-malignant tissue samples, the directionality of the line being much greater than for CB\(_1\)R → pEGFR in the bootstrap analyses (Fig. 2A). An edge LRIG1 → pEGFR was also found for the

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**Fig. 1.** Bivariate correlations between CB\(_1\) receptor scores and other parameters in the database. Shown are the Spearman rho values and the 95% confidence limits. Values where the Spearman rho value was >0.2 are shaded gray. Note that for the non-malignant scores, FAAH and pEGFR are available for both basal and luminal epithelial cells. The luminal scores were used here. Note also that the sample size for non-phosphorylated EGFR is much smaller than for pEGFR and most of the other parameters, thereby precluding its use in the subsequent Bayesian network analyses.
complete dataset, but this was deemed to be unreliable in the bootstrap analysis, since its frequency was lower than that for no preference. The addition of FAAH to the dataset did not change the observed pattern

\[ \text{pEGFR} \rightarrow \text{CB1R} \]

and suggested an additional edge \( \text{pEGFR} \rightarrow \text{FAAH} \) (Fig. 2B).

For the tumor samples, the networks obtained for a three-variable analysis (CB1R, pEGFR, and ErbB2, i.e., the three receptors in the sample), two four-variable analyses (the receptors and LRIG1; the receptors and FAAH), and a five-variable analysis (the receptors, LRIG1 and FAAH) are shown in Figure 3 for the whole patient sample ("original data") and the bootstrap analyses shown in Figure 4. In this case, the observed direction of the edges was highly dependent upon the inclusion, or not, of FAAH. When FAAH was excluded from the analyses, the directionality of the edges was from CB1R to the growth factor receptors and not vice versa (Fig. 3A, B), whereas when FAAH was included as a parameter, the directionality of the association between pEGFR and CB1R was reversed, that is, to be the same as seen in the non-malignant tissue (Fig. 3C, D).

Two other edges, LRIG1 \rightarrow \text{ErbB2} and \text{ErbB2} \rightarrow \text{FAAH} were seen in the tumor samples (Fig. 3B–D). There is some dispute as to the level of expression of ErbB2 in Pca, not the least due to the use of different antibodies [38]. In consequence, we reanalyzed the tumor data excluding this parameter, and found that for both CB1R, pEGFR and LRIG1 as variables (n = 271), and for CB1R, pEGFR, LRIG1, and FAAH as variables (n = 268), the directed acyclical graphs gave the edge pEGFR \rightarrow CB1R with a frequency of 65% and 62%, respectively, in the bootstrap analyses (data not shown).

A high CB1 receptor expression per se is insufficient to confer a selective survival of Pca cells to CB1 receptor stimulation.

The network analysis identifies CB1 receptors as down-stream of the EGFR, but does not provide any information as to whether a high CB1 receptor expression directly biases tumor cells towards survival in the presence of a CB receptor agonist, such as has been seen in astrocytoma clones transfected with this receptor [34]. In order to shed light on this possibility, AT1
prostate cancer cells were transfected with either eGFP or CB1/eGFP plasmids (the same plasmids used in [34]). This cell line is one of the R3327 Dunning rat prostatic tumor sublines, which show different metastatic abilities when injected into syngenic Copenhagen rats [39]. It has been used in vivo in orthotopic studies to characterize tumor growth in the ventral prostate after different interventions [40,41], and is thus a rather useful rodent cell line. We have previously characterized it with respect to its ability to accumulate and metabolize endocannabinoids [42,43], but to our knowledge the effect of CB1 receptor activation upon AT-1 cell survival has not been studied.

Successful transfection with the receptor was confirmed by RT-PCR, where the level of mRNA signal for the CB1 receptor was in the ratio 1: 2.6: 750 for untransfected, eGFP- and CB1/eGFP-transfected cell populations (data not shown). Rather than select individual clones with different receptor expression levels, we elected to utilize the variation in transfection efficiency of the unselected cells to mimic the large variation in CB1 receptor expression seen in the Pca tumor arrays [19]. In the cells transfected with the reporter protein eGFP alone, the CB receptor agonist CP55,940 produced a weak mitogenic effect, whereas in the cells transfected with the plasmid encoding CB1 receptors and eGFP, nanomolar concentrations of CP55,940 reduced the cell density (Fig. 5). Thus, introduction of the CB1 receptor into the cells conferred a changed sensitivity to CP55,940. Notably, the sensitivity to this agonist was the same for cells with expression levels above the median for the cell population as for those with expression levels below the median (Fig. 5). This result suggests that in the model system used, cells expressing levels of CB1 receptor that are above the median value do not exhibit an advantage in terms of cell proliferation and resistance to the anti-proliferative effects of CP55,940.

**DISCUSSION**

In the present study, Bayesian network analyses were undertaken on Pca tissue microarray data to identify potential upstream regulators of CB1 receptors. The strength of the study lies in the nature of the samples used, in that they were taken prior to diagnosis, and thus the parameters under study are not influenced by anti-cancer treatment of the patients. Further, the Bayesian methodology has proven useful in identifying potential biological networks [21,22]. This notwithstanding, at the outset, several potential limitations to the approach used in the present study should be considered. First, the model used assumes the data are continuous, whereas in fact they are
Fig. 5. Effect of the CB receptor agonist CP55,940 upon the density of AT1 cells transfected with CB1 receptors. Panel A shows a representative FACS for AT1 cells transfected with the CB1/eGFP plasmid and treated for 3 days with 0, 10, or 100 nM CP55,940. Panel B shows the number of cells and Panel C the % of cells above the cut-off chosen (shown as a dashed horizontal line marked with the arrow in Panel A) following CP55,940 treatment. Panels B and C show means and s.e.m. values, n = 6. The number n refers to the number of series run for FACS from each six well plate; in some experiments two series were run on the same day. In Panel A, a two-way ANOVA for repeated measures (matching for CP55,940) gave a significant interaction (F2,20 = 24.86, P < 0.0001). **P < 0.01, ***P < 0.001, Sidak’s multiple comparison test versus the corresponding value in the absence of CP55,940. In Panel C, there was no significant interaction (F2,20 = 1.56, P = 0.23) or main effect of CP55,940 (F2,20 = 1.50, P = 0.25) although there was a significant effect of the transfection with respect to the proportion of cells above the cut-off for the eGFP-transfected cells than for the CB1/eGFP-transfected cells (P1,10 = 159, P < 0.0001).

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Fig. 4. Bootstrap analyses of the Bayesian network inference analyses shown in Figure 3. The graphs show the percentage of times, out of 1,000 trials, that the directions A → B (dark grey), B → A (light grey), or no preference (white) was found. The graphs are: top left, CB1R, pEGFR, and ErbB2; top right, LRIG1, CB1R, pEGFR, and ErbB2; bottom right, FAAH, CB1R, pEGFR, and ErbB2; bottom left, FAAH, LRIG1, CB1R, pEGFR, and ErbB2. The dotted line is set at 50% to aid the reader.

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the CB1 receptor expression was down-stream of
in relevant biological systems. This light, they do form the basis for hypothesis testing
without issues and the results should be considered in
mice [33]. Thus, although the methodology is not
difference in
was seen, but only when FAAH was included. The
knowledge, EGFR- and ErbB2-receptor mediated regu-
luminal tissue [11]. A second novel observation was
in the tumor samples than in the non-malignant
malignant and tumor sample is in addition to be
considered. In addition to identifying potential upstream modulators of endocannabinoid signaling, the network analysis can provide conceptual data for formulation of novel hypotheses. In the present context, it allows us to suggest a way in which the CB1 receptor is involved in the pathogenesis of Pca. Our working model is based on the network analysis, the study of CB1 agonist effects upon EGF-stimulated Pca cells [15], the observations of down-stream activation of CB1 [18,19], and the transfected AT1 cell data presented here which suggest that a high CB1 receptor expression alone is not sufficient to drive the cells towards a survival phenotype in the presence of a CB receptor agonist. The model is summarized in Figure 6. In non-malignant tissue, normal EGF signaling produces both an array of cellular responses [51] and increases CB1 receptor expression. These then respond to the circulating levels of endocannabinoids and decrease EGFR expression [15], thus operating as a negative feedback regulatory mechanism. The findings that the 2-AG metabolizing enzyme acylglycerol kinase is expressed in Pca tumors to a higher level than in matched non-malignant tissue, and its down-regulation inhibits the motility of Pca cells in response to EGF [13] also support this model. In the tumor tissue, EGF-mediated signaling is overactive [27,52] and FAAH, acylglycerol kinase and other enzymes known to metabolize endocannabinoids (such as cyclooxygenase-2) are up-regulated [10–13], in the case of FAAH perhaps due in part to aberrant ErbB2 signaling that has been reported in some Pca cases [38,53]. This would result in a reduction in the levels of circulating endocannabinoids, thus weakening the feedback regulatory pathway.

It should be stressed that the model as shown in Figure 6 is theoretical rather that proven, although supported by current data. Clearly, it is important to determine in experimental models, cell cultures and tumor samples whether the model is valid. Additionally, given that a high tumor CB1 receptor expression is

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also associated with a poor prognosis in pancreatic cancer and stage II microsatellite-stable colorectal cancer [54,55] (but not hepatocellular carcinomas, [56]), and with the severity of ovarian cancer [57], our study raises the question as to whether similar networks are operative in these systems.

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